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High-throughput phenotyping using aerial images for predicting agronomic traits in soybean breeding programs

Pérez, O. M. 1

¹Instituto Nacional de Investigación Agropecuaria (INIA), Sistema Agrícola-Ganadero, Área de Ecofisiología y Manejo de Cultivos, Estación experimental La Estanzuela, Uruguay ROR

Editor

Esteban Casaretto

Universidad de la

República, Montevideo,

Uruguay

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☐ Correspondence

Osvaldo M. Pérez operez@inia.org.uy

Abstract

Plant breeding programs know the advantages of high-throughput phenotyping (HTP) in increasing efficiency over classical phenotyping and screening methods, which is achieved by saving time and improving selection accuracy. Even so, most programs have not yet systematically implemented this technology into their breeding pipelines. This review aims to indicate the restrictions of implementing HTP at a large scale and to summarize studies according to the used devices, data classes collected, and artificial intelligence (AI) methods applied to predict and classify agronomic traits in plant breeding programs with a focus on soybean [Glycine max (L.) Merr.]. Excluding HTP platforms in laboratories and greenhouses, satellite remote sensing, and autonomous mobile robots, this review focuses on field-based HTP platforms that take aerial images from drones and apply AI methods to associate those images with the traits of interest. Field-based HTP research is also conducted using hand-held devices that record individual vegetation indices (e.g., NDVI), a few spectral bands (multispectral radiometers), or the continuous range of the electromagnetic light spectrum (spectroradiometers). However, plant breeders must evaluate thousands of experimental lines each year, so using these devices instead of drones implies a trade-off between acquisition accuracy and the time it takes to collect the data. A challenge in the coming years is fine-tuning scalable, reliable models and optimizing data input, processing, and output pipelines to provide breeders with helpful information before they make selec-

Keywords: digital agriculture, machine learning, phenomics, remote sensing, soybean

Fenotipado de alto rendimiento usando imágenes aéreas para predecir rasgos agronómicos en programas de mejoramiento de soja

Resumen

Los programas de fitomejoramiento conocen las ventajas del fenotipado de alto rendimiento (FAR) para incrementar la eficiencia de los métodos de fenotipado clásico y cribado, lo cual se logra ahorrando tiempo y mejorando la precisión de selección. Aun así, la mayoría de los programas todavía no han implementado sistemáticamente esta tecnología en sus líneas de mejoramiento. Esta revisión tiene por objetivo indicar restricciones de implementar FAR a gran escala y resumir estudios según dispositivos usados, clases de datos recolectados y métodos de inteligencia artificial (IA) aplicados para predecir y clasificar caracteres agronómicos en programas de mejoramiento vegetal con foco en soja [*Glycine max* (L.) Merr.]. Excluyendo plataformas FAR en laboratorios e invernáculos, sensoramiento remoto satelital y robots móviles autónomos, esta revisión se enfoca en plataformas FAR a nivel de campo que toman imágenes aéreas desde drones y



aplican métodos de IA para asociar esas imágenes con caracteres de interés. Investigaciones en FAR a nivel de campo también se realizan con dispositivos portátiles que registran índices de vegetación individuales (por ejemplo, NDVI), unas pocas bandas espectrales (radiómetros multiespectrales) o el rango continuo del espectro electromagnético de la luz (espectrorradiómetros). Sin embargo, los fitomejoradores deben evaluar miles de líneas experimentales cada año, por lo que usar estos dispositivos en vez de drones implica un compromiso entre precisión de adquisición y el tiempo que lleva colectar los datos. Un desafío en los próximos años es ajustar modelos escalables y confiables y optimizar canales de entrada, procesamiento y salida de datos para proveer información útil a los mejoradores antes de realizar las selecciones.

Palabras clave: agricultura digital, aprendizaje automático, fenómica, sensoramiento remoto, soja

Fenotipagem de alto rendimento usando imagens aéreas para prever características agronômicas em programas de melhoramento de soja

Resumo

Os programas de melhoramento de plantas conhecem as vantagens da fenotipagem de alto rendimento (FAR) para aumentar a eficiência dos métodos de fenotipagem clássico e triagem, o que é alcançado economizando tempo e melhorando a precisão da seleção. No entanto, a maioria dos programas ainda não utiliza sistematicamente essa tecnologia em suas linhas de melhoramento. Esta revisão tem como objetivo indicar as restrições de implementar FAR em larga escala e resumir estudos sobre dispositivos utilizados, tipos de dados coletados e métodos de inteligência artificial (IA) aplicados para prever e classificar características agronômicas em programas de melhoramento de plantas com foco na soja [Glycine max (L.) Merr.]. Excluindo plataformas FAR em laboratórios e casas de vegetação, sensoriamento remoto via satélite e robôs móveis autônomos, esta revisão se concentra em plataformas FAR-de-campo que capturam imagens aéreas por drones e aplicam métodos de IA para associar essas imagens a características de interesse. A pesquisa FARde-campo também é conduzida com dispositivos portáteis que registram índices de vegetação individuais (e.g., NDVI), algumas bandas espectrais (radiômetros multiespectrais) ou o espectro eletromagnético contínuo da luz (espectrorradiômetros). No entanto, os fitomelhoristas devem avaliar milhares de linhagens experimentais a cada ano, pelo que a utilização destes dispositivos em vez de drones implica um obstáculo entre a precisão da aquisição e o tempo necessário para a obtenção de dados. Um desafio nos próximos anos será ajustar modelos escaláveis e confiáveis e otimizar canais de entrada, processamento e saída de dados para fornecer informações úteis aos melhoristas antes que façam as seleções.

Palavras-chave: agricultura digital, aprendizado automatico, fenômica, sensoriamento remoto, soja

1. Introduction

Breeding programs have only recently started using high-throughput phenotyping (HTP) platforms for predicting agronomic traits such as biotic and abiotic stress, date of physiological maturity, lodging, plant biomass, and grain yield⁽¹⁾. As imagery obtained by remote sensing has become readily available and affordable, efforts to use this technology have been on the rise to increase the genetic gain per unit of time. Although the traits mentioned above and others have already been subject to predictive and classification studies applying HTP, as in the case of maize [Zea mays L.], wheat [Triticum aestivum L.], rice [Oryza sativa L.], and soybean [Glycine max (L.) Merr.], among other crops, implementing these methods in breeding programs has been difficult⁽¹⁾⁽²⁾. This difficulty is mainly due to the uncertainty about the validation and repeatability of the predictions and the challenges in processing the data in a short time when data science is still an emerging discipline. This uncertainty refers to the fact that many HTP studies validate their models in independent data subsets of the same experiment from where they were trained but do not test them in independent environments. Even when using several experiments for model training, they may not repeat accurate predictions to the extent that the same genotype can express different phenotypes across locations and years due to the environmental conditions, and mainly if multiple genes control the predicted trait⁽³⁾. Data processing is more challenging in breeding programs



of developing countries than in private corporations, international programs, and public programs from developed countries because the latter have larger budgets and make rapid assimilation of the cute-edge technologies associated with earlier education of new users of these technologies.

Besides facilitating the classification and prediction of agronomic traits, applying HTP can help identify genomic regions associated with traits of interest. This can be done by combining phenomics with other omics approaches, such as genomics, which have a high potential to advance genetic gain in plant breeding⁽⁴⁾⁽⁵⁾. For example, Tanger and others⁽⁶⁾ performed a genotyping-by-sequencing analysis on a rice population of 1,516 recombinant inbred lines, constructed a genetic map, and performed a QTL (quantitative trait locus) mapping. However, they used a field-based HTP platform instead of the classic field phenotyping. As a result, they mapped genomic regions associated with four alleles that would have a negative effect on grain yield.

This review focuses on field-based HTP platforms that take aerial images from unmanned aerial vehicles (UAV) and apply artificial intelligence (AI) methods to associate the imagery with traits of interest in soybean plant breeding. By using AI to analyze multi-dimensional data sets collected with this technology based on remote sensing and geographic information systems (GIS), breeders could have helpful information to make better decisions when they select the best experimental lines. The studies and other references included here have been cited in peer-reviewed articles, provide a framework for discussing insights into implementing HTP at a large scale, and summarize predictive results according to the devices used, AI methods applied, and data classes collected in single or multiple environments. An extensive review is presented only for studies predicting physiological maturity in soybean. Several relevant studies for other traits and crops are not included because the number of articles published on field-based HTP is already in the hundreds. Some studies based on hand-held devices are also mentioned because the data acquisition in the field is easy or because accurate canopy measurements can be collected (e.g., spectroradiometers). Studies conducted with HTP platforms in laboratories and greenhouses, using tripods or towers –based or not on satellite remote sensing and GIS–, and autonomous mobile robots that use these technologies are not included.

2. What HTP Is, What the HTP Platforms Are, and What the Difference Is with Phenomics

HTP allows researchers and breeders to collect phenotypic data at a larger scale and in a shorter time than classic phenotyping, which comprises taking notes by visual appreciation or using measuring instruments with slow data acquisition. One of the pioneering studies applying HTP in plant sciences was conducted by Boyes and others⁽⁷⁾. By studying plant growth and development over the life cycle of Arabidopsis mutant lines with defects in specific biochemical pathways, these authors identified subtle phenotypic differences that had not been identified before using only classic phenotyping. Thus, compared to classic phenotyping, the HTP efficiency can be greater mainly when several traits are monitored simultaneously, but also the effectiveness, as happened with the study of Boyes and others⁽⁷⁾ identifying slight phenotypic differences due to genetic variation and environmental stress. Another advantage related to this is that by applying HTP researchers and breeders can collect information without destroying the experimental unit, which is often the case in classic phenotyping (e.g. when assessing growth rate).

A HTP platform in plant sciences is a system that integrates the use of devices and sensors, automated and eventually robotic procedures to collect the data, specific software for data extraction and analysis, and high computational resources to run and save big data with this software (i.e., workstations, local servers, or cloud servers). HTP platforms can be divided into two main groups: 1) laboratory and greenhouse HTP platforms and 2) field-based HTP platforms. The first studies applying HTP started in laboratory platforms, studying behavioral mutations in mice⁽⁸⁾ and later in plant sciences, as the study mentioned above in Arabidopsis⁽⁷⁾. It is in

laboratories and greenhouses that interdisciplinary teams performing HTP using state-of-the-art facilities can achieve the highest level of precision in plant-based phenotyping. Meanwhile, field-based HTP platforms include the ones using hand-held devices and those collecting multi-dimensional data sets based on remote sensing and GIS.

The concepts of phenomics and HTP are often interchanged; however, Araus & Cairns⁽⁹⁾ illustrated that the phenomics scope goes further. While HTP is the art of phenotyping a large number of accessions with greater efficiency in a non-destructive manner, phenomics is 'the acquisition of high-dimensional phenotypic data on an organism-wide scale' that allow a better understanding of the pathways linking genes to traits⁽¹⁰⁾. A few interdisciplinary research centers distributed worldwide have the facilities and high-tech HTP platforms to apply phenomics at the highest evaluative comprehension level. Several of these research centers applying phenomics are associated with the International Plant Phenotyping Network (https://www.plant-phenotyping.org/). Even though they may focus on conducting plant-based research (i.e., plants contained in plots), their scope reaches field-based HTP studies whose results can be applied more readily to plant breeding.

Pipeline workflows applied by some plant-based HTP platforms have been packed as technological products, such as PhenoTrack3D, which was developed to track maize organs over time⁽¹¹⁾. Several other technological products are available for plant-based HTP, such as those reviewed by Gill and others⁽¹²⁾ for plant stress phenotyping. However, many other HTP platforms have been individually customized by each interdisciplinary team according to the crop, the trait under study, the resources, and the know-how each team has. Interdisciplinary teams and customized pipelines were the standard when the first HTP platforms started to be developed, as is the case of Xiang & Tian⁽¹³⁾, who even had to program the missions to fly an autonomous helicopter. This pipeline step for planning flight missions is now fully automated by the specialized software that comes with the drone kit. Users now only must choose flight height, the overlapping percentage among images to both sides of the experiment, or how to fly over the experiment (e.g., in a zigzag design). Similarly, other pipeline steps that include complex codes are constantly summarized in functions included in packages that are uploaded, for example, to the 'R Project for Statistical Computing' (https://www.r-project.org) or to 'GitHub' (https://github.com), an Al-powered developer platform. A customized pipeline example, applied in the University of Illinois at Urbana-Champaign soybean breeding program, is shown for predicting the date the plant rows reach physiological maturity (Figure 1).

3. Field-Based HTP Platforms: Devices, Data Class Collected, and Artificial Intelligence (AI) Methods Used in Studies Conducted on Soybean and Other Crops

Multi-rotors are the most common UAV used to image field experiments as they are more affordable compared with fixed-wing drones or other rotary drones (helicopters). Typically, cheap UAV carry digital RGB (red, green, blue) cameras, while professionals often integrate more expensive multispectral or hyperspectral cameras. Although multispectral cameras may have a lower resolution than RGB cameras, a significant advantage of the former is that their lenses can record images at frequencies beyond the spectrum visible to the human eye, such as the red edge and near-infrared (NIR) spectrum bands. Both bands are the most commonly recorded by multispectral cameras, besides the RGB bands, and can be used to calculate several of the vegetation indices used in agricultural sciences. For example, the normalized difference vegetation index (NDVI = [NIR – red] / [NIR + red]) (Kriegler and others cited by Huang and others⁽¹⁴⁾), and the normalized difference red edge (NDRE = [NIR – red edge]) / [NIR + red edge]) (INIR + red edge]) (INIR + red edge]) (INIR + red edge]) (INIR + red edge])



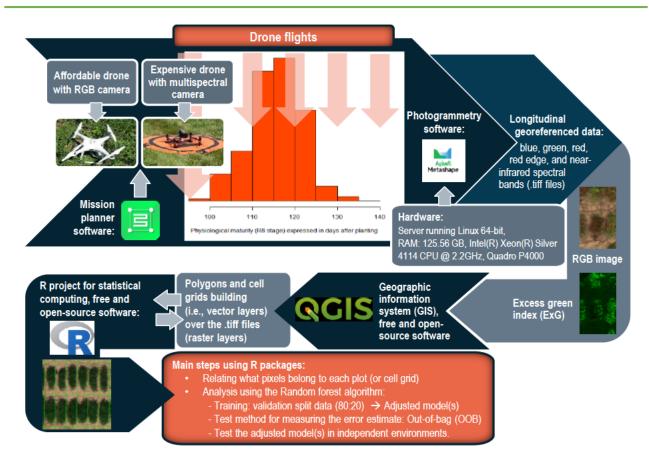


Figure 1. Pipeline workflow diagram of a high-throughput phenotyping platform applied in the University of Illinois at Urbana-Champaign soybean breeding program to predict the date the plant rows experiments reach physiological maturity (R8 stage)

The histogram shows the distribution of R8, expressed in days after planting (May 22, 2018), for the number of plant rows used to train the model (n = 7403).

Both reflectance indices, NDVI and NDRE (also called red edge NDVI, RENDVI), have been correlated with important agronomic traits such as plant growth, leaf area index, aboveground biomass, grain yield, flowering time, nitrogen status, and stress response⁽⁶⁾⁽¹⁶⁾⁽¹⁷⁾. These are only two of more than one hundred vegetation indices that can be calculated using multispectral imagery⁽¹⁷⁾. Multispectral band values can be recorded with multispectral cameras but also with hand-held multispectral radiometers. Using a multispectral radiometer and applying the power function regression model to analyze NDVI values, Ma and others⁽¹⁸⁾ explained the grain yield variation among 42 historical soybean cultivars (R^2 ranged from 0.44 to 0.80). In addition, hyperspectral cameras and hand-held spectroradiometers can also record multispectral imagery. The advantage of hyperspectral devices is that the continuous range of the electromagnetic light spectrum is divided into a much greater number of bands. This means that leaf and canopy attributes can be associated with a much greater number of reflectance indices⁽¹⁶⁾⁽¹⁹⁾⁽²⁰⁾⁽²¹⁾.

Another sensor that has applications in plant breeding is the thermal sensor, which is often built-in for hand-held devices and, more recently, multispectral cameras mounted on drones. Instead of capturing the scene's reflectance as the lenses recording the spectral bands do, the thermal sensors capture the infrared radiation emitted by objects in the field (i.e., the crop, weeds, and the ground). Thermal sensors detect changes in plant temperature mainly due to the trade-off between the opening and closing of stomata in response to the transpiration rate, water content, and stress signals, which include diseases such as the sudden death syndrome in soybean caused by *Fusarium virguliforme*(22)(23)(24). A couple of limitations of the thermal sensors are that they are

expensive and have a lower spatial resolution than those achieved by lenses recording the spectral of the visible light and beyond, such as the red-edge and NIR spectrums⁽²⁵⁾. Thus, in plant breeding, researchers must reconcile the spatial pixel resolution of the thermal sensor with the plot size of the breeding experiments. For example, Kaler and others⁽²⁶⁾ conducted an association mapping study with 345 soybean accessions to identify loci for canopy temperature under drought conditions. Instead of the classic phenotyping, the authors used a field-based HTP platform with a thermal infrared camera (resolution 640 × 512 pixels) mounted on a tethered balloon filled with helium and held at 75 m. In this case, the resolution was enough to associate the pixel information to the corresponding size plot, 3.65 m in length with 0.76 m row spacing (two-row plots) in one experiment, and 4.57 m in length with 0.19 m row spacing (seven-row plots) in the other experiment.

Plant breeders are also interested in predicting physiological maturity using remote sensing, mainly in plant-row trials because of the large amount of time required to take notes for several thousand experimental lines. In the case of soybean, physiological maturity is the R8 growth stage⁽²⁷⁾, defined as when 95% of pods reach their mature color, and the maximum biomass accumulation in the seed occurs. As a field-based HTP case study, the studies conducted to date in single and multiple environments using different methods of analyzing RGB and multispectral images to predict soybean physiological maturity are listed (**Table 1**). Using the machine learning algorithm Random forest (RF) and a binary prediction model applied to multispectral aerial images taken over plant rows at the University of Illinois, Yu and others⁽²⁸⁾ reported an overall accuracy of 93.8% for classifying R8. The authors also remotely measured the canopy area and the length of the plant rows and reported correlations with grain yield of r = 0.56 and r = 0.49, respectively. By using RGB images and deep convolutional neural networks (CNN), Trevisan and others⁽²⁹⁾ and Moeinizade and others⁽³⁰⁾ also trained models to predict the R8 stage in soybean. In several trials evaluated in different environments, both studies reported a root mean squared error (RMSE) of approximately 2 days for the average of analyses, a tolerable value considering that breeders most often take notes every 5-7 days. Still, these and other authors reported lower prediction errors for some experiments (**Table 1**).

Additional studies to predict physiological maturity in soybean have been conducted using methods different from RF and CNN (Table 1). Using a ground-based field spectroradiometer to measure the canopy reflectance, Christenson and others⁽¹⁹⁾ applied partial least squares regression (PLSR) to associate the R8 stage with 91 wavebands and four vegetation indices with different versions for its calculation. These were six versions for RENDVI, the blue, green, and red NDVIs, and three for the water index. After the PLSR analysis, one model was adjusted with the most significant indices and the other with the most significant individual wavebands (RMSE = 5.5 and 5.2 days, respectively). A possible reason to explain the fact that these prediction errors were more than twice the errors obtained by the last two studies mentioned above⁽²⁹⁾⁽³⁰⁾ is the *n*-value used to train the models. While Christenson and others⁽¹⁹⁾ used 40 cultivars, the other two studies used hundreds to several thousands of experimental lines. It is noteworthy that with a ground-based field spectroradiometer, it is possible to collect a huge amount of spectral information with high accuracy. However, the number of plots an individual can phenotype daily is a few hundred, while a drone carrying a camera can phenotype ~10,000 plant rows in half an hour.

Applying PLSR to multispectral images taken from a drone to 326 soybean lines, Zhou and others⁽³³⁾ explained up to 70% of the R8 stage variation (RMSE = 1.7 days). This best result occurred using images from the second of three flight dates, five linear components in the PLSR model, and the rating change of image features (from the second to the third flight) as predictors. In turn, the above results were improved by adjusting the maturity records annotated in the field with the variances of two canopy image features (red edge NDVI and canopy chlorophyll content index) obtained from soybean lines that matured the same day according to the predictions (**Table 1**). However, applying the same adjustment technique but using images of the third flight instead of the second flight, the accuracy of the maturity predictions did not improve compared to the same analysis but without adjusting the records annotated in the field ($R^2 = 0.71$ and RMSE = 1.6 days).



Table 1. Field-based high-throughput phenotyping studies conducted in single and multiple environments using different methods of analyzing RGB (red, green, blue) and multispectral images to predict physiological maturity in soybean

Device	Spectral bands	Vegetation indices (VIs)	Tested in independent ent envi-	Envi- ron- ments	Loca- tions	Years	Total assessed field data (n)	Analytical models	Maturity assess- ment approach	Indicators according to the data class considered (numerical or categorical)					
										RMSE (days)	R ²	r	Карра	Overall accuracy	Reference
Spectro- radiometer	550, 680, 715, 915, 990, & 1140 (nm)	-		3	3	2	240	PLSR (partial least square regression)	Maturity date	5.2	0.50	-	-	-	Christenson and others ⁽¹⁹⁾
UAV	RGB (red, green, blue) & near-infrared (NIR)	7		1 (two trials)	1	1	3220	Random forest (RF)	Mature & immature for a given date (i.e., nearest temporal image data)	-	-	-	0.73	93.8%	Yu and others ⁽²⁸⁾
	RGB	Normalized green excess index (NGE)		22	22	1	Data shown in a histo- gram	Piecewise linear regression model	Maturity date	-	-	0.92	-	-	Narayanan and others ⁽³¹⁾
Manually operated cart	NOD	20		4	4	1	6039	PLSR, RF, Cubist, Artificial neural networks (ANN), & Support vector machine (SVM)	Days to maturity after planting	3.7	0.76	-	-	-	Yuan and others ⁽³²⁾
UAV	Multispectral (red, green, blue, red edge, &NIR)	20		1	1	1	326	PLSR	· Maturity date	1.4	0.81	-	-	-	Zhou and others ⁽³³⁾
Hand-held sensor	Red (660 nm) & NIR (780 nm)	NDVI		4	2	2	96	Inverse logistic model		1.3	0.92	-	-	-	Lindsey and others ⁽³⁴⁾
	RGB	-	Yes	5	5	2	15622	Convolutional neural networks (CNN)		1.4	-	-	-	-	Trevisan and others ⁽²⁹⁾
UAV	Multispectral	Several, but the final model was adjusted only with the spectral bands		4	3	3	1048	Deep learning, RF, SVM, & Linear regression	Days to maturity after emergence	8.0	-	0.66	-	-	Teodoro and others ⁽³⁵⁾
	RGB	Greenness leaf index (GLI)	No _	5 (53 trials)	3	2	4415	Nonparametric local polynomial regression (LOESS)	- Maturity date	1.7	0.95	0.97	-	-	Volpato and others ⁽³⁶⁾
				6	4	2	6350	CNN-Long short-term memory & LOESS		0.9	0.95	-	-	-	Moeinizade and others ⁽³⁰⁾
		Leaf chlorophyll content & fractional vegetation cover		1	1	1	153	PLSR, multiple stepwise regression, RF, & Gaussian process regression	Mature & Immature	-	-	-	-	98.8%	Hu and others(37)
		-	No but in a nearby trial	1 (two trials)	1	1	2662	DS-Soybean Net (a CNN method)	Immature, near- mature, mature, & harvested	-	-	-	-	86.3%	Zhang and others ⁽³⁸⁾
	RGB & multispectral	Excess green index (ExG)	Yes	3	1	3	32191	RF (binary & regression predictions)	Maturity date	1.3	0.91	-	0.35	40.2%	Pérez and others(39)

The indicators show the results of those experiments and analyses that obtained the lowest prediction error.

Narayanan and others⁽³¹⁾ calculated a normalized green excess index and adjusted a piecewise linear regression model in function of time. For a subset of test data and locations, the adjusted models showed a range in the Pearson correlation coefficient (*r*) from 0.79 to 0.92 (**Table 1**). Based on these results, Volpato and others⁽³⁶⁾ conducted a comparative analysis, deciding after testing two other vegetation indices to use the same normalized green excess index with a nonparametric local polynomial regression model. Finally, one of the most extensive studies on soybean was conducted by Yuan and others⁽³²⁾, who used RGB images and five regression and classification models to predict the date of R8, plant height, seed size, protein, oil, fiber, and grain yield. In the case of physiological maturity, the models explained up to 76% of the R8 stage variation but with a higher error than the other studies cited previously (RMSE = 3.7 days).

Trevisan and others⁽²⁹⁾ and Moeinizade and others⁽³⁰⁾ stated that an advantage of CNN is that they benefit from the spatial structure of the pixels. This advantage could be especially important when CNN is applied to complex traits, such as plant development (e.g., morphology, plant growth, and plant/organ counting) and plant stress⁽⁴⁰⁾. Physiological maturity is a trait strongly associated with the loss of green in the leaves; therefore, predicting maturity using this easy-to-observe change in color is one reason why Moeinizade and others⁽³⁰⁾ achieved good performance with CNN after using just four convolutional layers. A convolutional layer comprises the training of specific details of images (features) that are automatically learned and stacked hierarchically on other layers. Other researchers have found that RF can also be applied to predict complex traits. For example, in a comparative analysis between three machine learning algorithms for wheat lodging classification, Zhang and others⁽⁴¹⁾ obtained lower overall accuracy fluctuations between and within ten replicates of datasets from three different dates when they used RF compared to neural networks or support vector machine (SVM). In the same work, when RF and the best of three deep learning methods –GoogLeNet, CNN, and VGG-16– were compared, no significant differences were identified (*P*<0.05).

Based on classification and regression trees (CART), RF is also a machine learning algorithm that ensembles trees (i.e., the predictors) by using the bootstrap aggregation method, also known as the bagging method (42)(43). Emulating branches in a tree, the splitting in each node is a sequential decision process, meaning that the splitting occurs from the base or root node to the top leaf; this is the terminal node where the splitting events can no longer happen. A difference from single decision trees is that the RF algorithm uses the bagging method, which implies that the splitting occurs based on a random selection of features with replacement from the original training set that can be categorical or numerical(42)(43). Even with the risk of high bias, the aim of training different random subsets within the training set (i.e., bagging) is to decrease the correlations between trees and the variance, which can cause overfitting of the trained model(44). Through learning algorithms associated with their respective bootstrap samples, the final ensemble of the trees results in a single classifier or regressor, according to the type of the dependent variable(43).

Pubescence color (gray, light tawny, and tawny) is an important soybean breeding trait that was recently classified using aerial images taken from a UAV⁽⁴⁵⁾. Breeders take notes for this qualitative trait to characterize the experimental lines and trace whether the identity in the field and the progenies' frequencies are according to the expected. Two loci epistatically control pubescence color, and the gene combinations that give the three phenotypes are tt__ (gray), T_tdtd (light tawny), and T_Td_ (tawny)⁽⁴⁶⁾⁽⁴⁷⁾. Applying the SVM algorithm to the multispectral images' analysis, Bruce and others⁽⁴⁵⁾ could classify gray and tawny with an overall accuracy in the tested data of 75% using the red/blue band ratio. However, they failed to separate light tawny from tawny pubescence because the algorithm incorrectly assigned all the light tawny lines as having tawny pubescence.

Lodging is another categorical and important trait in soybean, but unlike pubescence color, which is a nominal categorical variable, lodging is mainly considered by breeders as an ordinal categorical variable. Usually, this is by visually assigning a number on a scale from 1 to 5 in the field, where the higher the number, the greater the percentage of prostrated plants. Based on this scale, Roth and others⁽⁴⁸⁾ discarded their intention of developing



a drone-based lodging index because they could not extract the time at which plant height drops. This was related to the fact that when they conducted the photogrammetric processing, the depth maps were sensitive to defoliation and color changes due to the senescence phase. However, using the same ordinal categorical scale but divided into four levels: non-lodging [1, 1.5], moderate lodging [2, 2.5], high lodging [3, 3.5], and severe lodging [4, 4.5, 5], Sarkar and others⁽⁴⁹⁾ could classify lodging with an overall accuracy of 96% by applying artificial neural network (ANN), though this was after using a data balancing method (smote-ENN) to deal with the unbalanced number of plots per lodging level. Previously, although predicting wheat lodging instead of soybean and using a binary categorical variable (lodging and non-lodging), Zhang and others⁽⁴¹⁾ achieved an overall accuracy of ~90% by applying RF, SVM, and three deep learning methods without preprocessing the data with a balancing method.

Machine learning and deep learning are the two branches of AI methods most commonly applied for data analysis collected using a HTP platform⁽⁵⁰⁾; however, robotics and computer vision are two other increasing AI methods. On the other hand, non-AI methods such as process-based models⁽⁴⁸⁾, geospatial analysis⁽⁵¹⁾, and statistical and mathematical modeling combining genomic data⁽⁵²⁾ have also been applied for plant breeding purposes based on the imagery collected with a HTP platform. Within each one of the above AI and non-AI methods, several promising approaches with potential use in plant breeding have been reported for predicting main agronomic traits using HTP field platforms. However, there is no agreement about what learning algorithms and other analytic methods best predict the different traits. Undoubtedly, the genetic diversity of the germplasm, the size of the training and validation datasets, the number of trials across the different environments, the number of flights, the time and weather conditions while the images are taken, and the different analysis pipelines applied affect the results expressed in terms of R^2 and RMSE when the variable is numerical.

Even using the same methods and information, the results may vary according to the data type researchers assume when studying the same trait. Although two of the studies mentioned above were independent of each other, one studying lodging on wheat and the other on soybean⁽⁴¹⁾⁽⁴⁸⁾, the first one considered the trait as a binary categorical variable (lodging and non-lodging), and the second as an ordinal categorical variable (1-5 scale). Another example is predicting soybean physiological maturity, where independent studies considered different data types. While Yu and others⁽²⁸⁾ and Hu and others⁽³⁷⁾ considered the trait as a binary categorical variable (mature and immature plots), Zhang and others⁽³⁸⁾ did it as a nominal categorical variable (immature, near-mature, mature, and harvested plots), and others as a numerical variable to predict the date the plant rows reached the R8 stage (Table 1). Besides differences in the prediction error values that may result, it is noteworthy that the kind of information the results provide to the breeders also matters. For example, in the case of soybean physiological maturity, binary models can be helpful mainly for classifying maturity groups because these models do not predict what date the event occurred (i.e., the R8 stage). Meanwhile, considering a time series of images and the variable as numerical, a regression analysis can determine the date on which the same event occurred; so, besides maturity groups, the model is also helpful in predicting the germplasm cycle length expressed in days.

Compared to pubescence color, lodging, physiological maturity, and other traits in soybean and other crops, more research studies have been conducted to predict aboveground biomass and its relationship with grain yield. In one interesting study conducted on soybean, Maimaitijiang and others⁽⁵³⁾ adjusted a model combining canopy spectral and volumetric information, which they called "vegetation index weighted canopy volume model (CVM_{VI})." The regression analysis between the canopy volume with aboveground biomass determined through destructive field sampling indicated that this proposed model (CVM_{VI}) had a similar coefficient of regression ($R^2 = 0.893$) as more complex models such as PLSR ($R^2 = 0.911$), or stepwise multilinear regression ($R^2 = 0.915$). The predictions were made by first estimating canopy volume by photogrammetry, a technique highly correlated with the aboveground biomass⁽⁵⁴⁾. The volumes were then weighted by a bulk density factor calculated with the green red ratio index (GRRI = green/red) of pixels, chosen among other vegetation indices

due to its best fit. Using data from the same experiment, Maimaitijiang and others⁽⁵⁵⁾ applied multimodal data fusion within a deep neural network framework and compared the results with other learning algorithms. The deep neural network algorithm explained a higher proportion of the variation in grain yield (72%) compared to RF (66%). However, when only the RGB bands were used as features, RF was the algorithm that best explained the grain yield variation.

Two of the most important traits in soybean and other crops, physiological maturity⁽²⁹⁾⁽³⁰⁾ and aboveground biomass and its relationship to grain yield⁽⁵³⁾⁽⁵⁵⁾, were predicted with a low error using only RGB images collected with drones carrying affordable digital cameras. Multispectral cameras are several times more expensive than digital RGB cameras, mainly if they include a thermal sensor. Among other traits, they have been used more for studying traits related to plant health and plant stress, such as diseases, pests, and water deficit in soybean⁽⁵⁶⁾⁽⁵⁷⁾⁽⁵⁸⁾. Concerning field spectroradiometers collecting hyperspectral data, they are suitable for studying complex traits such as photosynthetic traits⁽¹⁹⁾⁽²⁰⁾⁽²¹⁾. However, there is a trade-off between the cost and benefit of including RGB, multispectral, or hyperspectral data for the predictive analyses.

Spectroradiometers are even much more expensive than drones carrying multispectral cameras. At the same time, spectroradiometers need more time to collect the same amount of data than flying a drone over the experiments. Therefore, collecting data with spectroradiometers would be a better option when RGB or multispectral data cannot provide proper or accurate information for certain traits. Otherwise, besides using more time than a drone carrying a camera (RGB or multispectral), redundant information may be collected with spectroradiometers, mainly when phenotyping simple traits. Still, this may also occur with complex traits when the relationships between light reflectance patterns and the underlying biological processes of plants are still poorly understood⁽¹²⁾, though first comprehensive studies are arising⁽²¹⁾. Phenotyping such a large amount of data is particularly challenging when plant breeders attempt to identify slight favorable genetic differences among thousands of experimental lines, commonly evaluated in small single-row plots. These considerations also apply to other self-pollinated crops besides soybean and cross-pollinated crops when selecting inbred lines based on hybrid performance.

4. Justification of Research Applying Field-Based HTP

Plant breeding programs of self-pollinated grain or legume crops, such as soybean, need to annually record phenotypic traits for thousands of experimental lines grown in plant rows to select those that should be evaluated in preliminary yield tests. Similarly, the same occurs when testing F₁ hybrids in cross-pollinated grain crops such as maize. In both cases, it is a time-consuming task that includes the characterization before harvest of the date to reach silking in maize or the R8 stage in soybean, only to mention a couple of traits. In the end, much of the data collected is not used because the vast majority of the plant rows or inbred lines in soybean and maize, respectively, will be discarded due to their low grain yield and not because of a too-short or too-long cycle length.

In the case of soybean, experimental lines are often developed from hybridizations between parents of different maturity groups. Hence, the range of days to reach the R8 stage in a breeding population can be wide between and within families, requiring a repeated collection of field notes every three or four days for about four to five weeks. In addition, because of the high number of lines to evaluate, breeding programs often conduct their selections in trials of one-row plots of plant rows. Therefore, the selection accuracy for grain yield can be compromised due to a lack of replication, but also because lines of different maturities can be affected differently by random weather conditions that occur each growing season.

The use of drones for HTP by taking aerial images across the growing season promises to improve the efficiency of breeding programs. This could increase efficiency by saving time or allow the evaluation of a greater number



of plots using similar resources, which at the same time increases selection intensity. In turn, by using the image features, phenotypic values could be estimated with higher accuracy than can be done by humans taking visual ratings in the field. For example, it was reported that for the R8 stage in soybean, predicted values were more reliable than ground-truth values⁽³⁶⁾. In this same crop, similar results could be obtained for lodging or pubescence color, considering that within a breeding program the notes are often taken by a team and not only by one person.

To date, only one study has been conducted for predicting pubescence color in soybean⁽⁴⁵⁾, while not many but several studies have predicted the R8 stage using different algorithms with aerial images or hyperspectral reflectance signatures of the canopy (**Table 1**). However, the small data set is a limitation of some studies, especially when using a spectroradiometer. Another relevant limitation is that only two of the studies predicting maturity were conducted across multiple environments and tested the models in an independent environment⁽²⁹⁾⁽³⁹⁾, highlighting the relevance of conducting more research to test the reproducibility of the models.

Although several studies have been conducted to predict aboveground biomass and grain yield, the same question remains about models' reproducibility because some studies have trained them using only a single environment and with low *n*-values. For instance, although Maimaitijiang and others⁽⁵³⁾⁽⁵⁵⁾ used the most cutting-edge algorithm, the study was conducted in a single environment with only three cultivars evaluated in multiple subplots. The uncertainty about the reproducibility of models is a concern in the breeders' community⁽²⁾. Poor reproducibility would occur mainly when complex traits are predicted using small datasets representing a unique environment or germplasm. In this sense, when RF was proposed, an advantage over other learning algorithms is that it includes the bagging method in the algorithm, a method that decreases the overfitting of the trained model⁽⁴⁴⁾. Concerning overfitting, models trained with shared check cultivars across environments had a better performance when tested in independent environments⁽²⁹⁾⁽³⁹⁾, and they were also more general when redundant image features were disregarded after conducting principal component analysis⁽³⁹⁾.

For complex traits such as plant growth, canopy volume, aboveground biomass, and grain yield, most predictive studies in soybean and other crops have obtained leaf or canopy reflectance records using only one kind of HTP platform. Three examples of this are Ma and others⁽¹⁸⁾, that used a portable multispectral radiometer; Yu and others⁽²⁸⁾, that used multispectral images; and Maimaitijiang and others⁽⁵³⁾⁽⁵⁵⁾, that used RGB images. At a breeding population scale, studies conducted in soybean relating canopy reflectance information obtained from different platforms or sources are absent⁽⁵⁹⁾⁽⁶⁰⁾. Instruments such as portable spectroradiometers have the advantage of recording the continuous range of the electromagnetic spectrum, which opens more opportunities to relate features to the traits of breeding interest⁽¹⁹⁾⁽²⁰⁾⁽²¹⁾. It is possible that more accurate predictions could be made for complex traits by associating features from time series of aerial images with hyperspectral reflectance signatures of the canopy.

5. Concluding Remarks

Several studies indicate that predicting traits of agronomic interest using a field-based HTP platform based on features of aerial imagery associated with the respective ground-truth values is possible. However, most studies applying HTP have validated their models in data subsets of the same environment where they were trained, which can lead to overfitting when the models are tested in a different environment. Overfitting may happen if too many image features or redundant information is used to train the models, the variation in the germplasm is narrow for the trait of interest, and the growing conditions among environments differ significantly. Considering this, collaborative international studies using the same workflow could fine-tune more generalized models by covering a broader spectrum of possible scenarios. Hence, to provide helpful information for breeders, there is

still a path to identify redundant information to adjust more general and reliable models when tested in new environments over the years or in different locations. More reliable predictions would increase the efficiency and accuracy of the methods for selecting the best lines, leading to breeding programs with higher genetic gains per breeding cycle.

Another critical challenge is processing and analyzing the data in a short time, before harvest or at least before breeders make the final selections in their attempt to choose experimental lines with a differential genetic value. Applying a HTP platform with immediate application for the plant breeding selection process requires developing a smooth pipeline workflow that requires investment not only in software and hardware but also in hiring or training collaborators with strong computational and analytical skills and a desirable background in agronomy, biology, genetics, or plant breeding. Data science is still an emerging discipline, though several universities and private companies have respectively created certificates, bachelor's and master's degrees, and training programs in the area. Merging all this knowledge in smooth pipeline workflows is still harder when HTP is an essential but not the only component of phenomics studies conducted to reveal the pathways between genes and traits. To the extent that the coming findings by applying HTP will be cooperative efforts integrated with other omics approaches, the larger will be the genetic progress of breeding programs.

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Transparency of data

Available data: The entire data set that supports the results of this study was published in the article itself.

Author contribution statement

	OM Pérez
Conceptualization	
Investigation	
Methodology	
Visualization	
Writing – original draft	
Writing – review and editing	

References

(1) Zhou J, Nguyen HT, editors. High-throughput crop phenotyping. Cham: Springer Nature; 2021. 249p. Doi: 10.1007/978-3-030-73734-4.



- (2) Araus JL, Kefauver SC, Zaman-Allah M, Olsen MS, Cairns JE. Translating high-throughput phenotyping into genetic gain. Trends Plant Sci. 2018;23(5):451-66. Doi: 10.1016/j.tplants.2018.02.001.
- (3) Poorter H, Hummel GM, Nagel KA, Fiorani F, von Gillhaussen P, Virnich O, Schurr U, Postma JA, van de Zedde R, Wiese-Klinkenberg A. Pitfalls and potential of high-throughput plant phenotyping platforms. Front Plant Sci. 2023;14:1233794. Doi: 10.3389/fpls.2023.1233794.
- (4) Edwards D, Batley J. Plant bioinformatics: from genome to phenome. Trends Biotechnol. 2004;22(5):232-7. Doi: 10.1016/j.tibtech.2004.03.002.
- (5) Deshmukh R, Sonah H, Patil G, Chen W, Prince S, Mutava R, Vuong T, Valliyodan B, Nguyen HT. Integrating omic approaches for abiotic stress tolerance in soybean. Front Plant Sci. 2014;5:00244. Doi: 10.3389/fpls.2014.00244.
- (6) Tanger P, Klassen S, Mojica JP, Lovell JT, Moyers BT, Baraoidan M, Naredo MEB, McNally KL, Poland J, Bush DR, Leung H, Leach JE, McKay JK. Field-based high throughput phenotyping rapidly identifies genomic regions controlling yield components in rice. Sci Rep. 2017;7:42839. Doi: 10.1038/srep42839.
- (7) Boyes DC, Zayed AM, Ascenzi R, McCaskill AJ, Hoffmann NE, Davis KR, Görlach J. Growth stage-based phenotypic analysis of Arabidopsis: a model for high throughput functional genomics in plants. Plant Cell. 2001;13(7):1499-510. Doi: 10.1105/tpc.13.7.1499.
- (8) Nolan PM, Kapfhamer D, Bućan M. Random mutagenesis screen for dominant behavioral mutations in mice. Methods. 1997;13(4):379-95. Doi: 10.1006/meth.1997.0545.
- (9) Araus JL, Cairns JE. Field high-throughput phenotyping: the new crop breeding frontier. Trends Plant Sci. 2014;19(1):52-61. Doi: 10.1016/j.tplants.2013.09.008.
- (10) Houle D, Govindaraju DR, Omholt S. Phenomics: the next challenge. Nat Rev Genet. 2010;11(12):855-66. Doi: 10.1038/nrg2897.
- (11) Daviet B, Fernandez R, Cabrera-Bosquet L, Pradal C, Fournier C. PhenoTrack3D: an automatic high-throughput phenotyping pipeline to track maize organs over time. Plant Methods. 2022;18(1):130. Doi: 10.1186/s13007-022-00961-4.
- (12) Gill T, Gill SK, Saini DK, Chopra Y, de Koff JP, Sandhu KS. A comprehensive review of high throughput phenotyping and machine learning for plant stress phenotyping. Phenomics. 2022;2(3):156-83. Doi: 10.1007/s43657-022-00048-z.
- (13) Xiang H, Tian L. Development of a low-cost agricultural remote sensing system based on an autonomous unmanned aerial vehicle (UAV). Biosyst Eng. 2011;108(2):174-90. Doi: 10.1016/j.biosystemseng.2010.11.010.
- (14) Huang S, Tang L, Hupy JP, Wang Y, Shao G. A commentary review on the use of normalized difference vegetation index (NDVI) in the era of popular remote sensing. J For Res. 2021;32(1):1-6. Doi: 10.1007/s11676-020-01155-1.
- (15) Gitelson A, Merzlyak MN. Spectral reflectance changes associated with autumn senescence of *Aesculus hippocastanum* L. and *Acer platanoides* L. leaves. Spectral features and relation to chlorophyll estimation. J Plant Physiol. 1994;143(3):286-92. Doi: 10.1016/S0176-1617(11)81633-0.
- (16) Peñuelas J, Filella L. Visible and near-infrared reflectance techniques for diagnosing plant physiological status. Trends PI Sci. 1998;3(4):151-6. Doi: 10.1016/S1360-1385(98)01213-8.
- (17) Xue J, Su B. Significant remote sensing vegetation indices: a review of developments and applications. J Sens. 2017;2017:1353691. Doi: 10.1155/2017/1353691.
- (18) Ma BL, Dwyer LM, Costa C, Cober ER, Morrison MJ. Early prediction of soybean yield from canopy reflectance measurements. Agron J. 2001;93(6):1227-34. Doi: 10.2134/agronj2001.1227.
- (19) Christenson BS, Schapaugh WT, An N, Price KP, Prasad V, Fritz AK. Predicting soybean relative maturity and seed yield using canopy reflectance. Crop Sci. 2016;56(2):625-43. Doi: 10.2135/cropsci2015.04.0237.

- (20) Meacham-Hensold K, Montes CM, Wu J, Guan K, Fu P, Ainsworth EA, Pederson T, Moore CE, Brown KL, Raines C, Bernacchi CJ. High-throughput field phenotyping using hyperspectral reflectance and partial least squares regression (PLSR) reveals genetic modifications to photosynthetic capacity. Remote Sens Environ. 2019;231:111176. Doi: 10.1016/j.rse.2019.04.029.
- (21) Montes CM, Fox C, Sanz-Sáez Á, Serbin SP, Kumagai E, Krause MD, Xavier A, Specht JE, Beavis WD, Bernacchi CJ, Diers BW, Ainsworth EA. High-throughput characterization, correlation, and mapping of leaf photosynthetic and functional traits in the soybean (*Glycine max*) nested association mapping population. Genetics. 2022;221(2). Doi: 10.1093/genetics/iyac065.
- (22) Ishimwe R, Abutaleb K, Ahmed F. Applications of thermal imaging in agriculture: a review. Adv Remote Sens. 2014;3(3):128-40. Doi: 10.4236/ars.2014.33011.
- (23) Kim M, Lee C, Hong S, Kim SL, Baek J-H, Kim K-H. High-throughput phenotyping methods for breeding drought-tolerant crops. Int J Mol Sci. 2021;22:8266. Doi: 10.3390/ijms22158266.
- (24) Hatton N, Sharda A, Schapaugh W, van der Merwe D. Remote thermal infrared imaging for rapid screening of sudden death syndrome in soybean. Comput Electron Agric. 2020;178:105738. Doi: 10.1016/j.compag.2020.105738.
- (25) Khanal S, Fulton J, Shearer S. An overview of current and potential applications of thermal remote sensing in precision agriculture. Comput Electron Agric. 2017;139:22-32. Doi: 10.1016/j.compag.2017.05.001.
- ⁽²⁶⁾ Kaler AS, Ray JD, Schapaugh WT, Asebedo AR, King CA, Gbur EE, Purcell LC. Association mapping identifies loci for canopy temperature under drought in diverse soybean genotypes. Euphytica. 2018;214:135. Doi: 10.1007/s10681-018-2215-2.
- (27) Fehr WR, Caviness CE. Stages of soybean development [Internet.] Ames: Iowa Agricultural and Home Economics Experiment Station Publications; 1977 [cited 2025 Aug 05]. 12p. Available from: http://lib.dr.iastate.edu/specialreports/87
- (28) Yu N, Li L, Schmitz N, Tian LF, Greenberg JA, Diers BW. Development of methods to improve soybean yield estimation and predict plant maturity with an unmanned aerial vehicle based platform. Remote Sens Environ. 2016;187:91-101. Doi: 10.1016/j.rse.2016.10.005.
- ⁽²⁹⁾ Trevisan R, Pérez O, Schmitz N, Diers B, Martin N. High-throughput phenotyping of soybean maturity using time series UAV imagery and convolutional neural networks. Remote Sens. 2020;12(21):3617. Doi: 10.3390/rs12213617.
- (30) Moeinizade S, Pham H, Han Y, Dobbels A, Hu G. An applied deep learning approach for estimating soybean relative maturity from UAV imagery to aid plant breeding decisions. Mach Learn Appl. 2022;7:100233. Doi: 10.1016/j.mlwa.2021.100233.
- (31) Narayanan B, Floyd B, Tu K, Ries L, Hausmann N. Improving soybean breeding using UAS measurements of physiological maturity. In: Thomasson JA, McKee M, Moorhead RJ, editors. Autonomous Air and Ground Sensing Systems for Agricultural Optimization and Phenotyping IV. Baltimore: SPIE; 2019. Doi: 10.1117/12.2519072.
- (32) Yuan W, Wijewardane NK, Jenkins S, Bai G, Ge Y, Graef GL. Early prediction of soybean traits through color and texture features of canopy RGB imagery. Sci Rep. 2019;9(1):1-17. Doi: 10.1038/s41598-019-50480-x.
- (33) Zhou J, Yungbluth D, Vong CN, Scaboo A, Zhou J. Estimation of the maturity date of soybean breeding lines using UAV-based multispectral imagery. Remote Sens. 2019;11(18):2075. Doi: 10.3390/rs11182075.
- (34) Lindsey AJ, Craft JC, Barker DJ. Modeling canopy senescence to calculate soybean maturity date using NDVI. Crop Sci. 2020;60(1):172-80. Doi: 10.1002/csc2.20079.
- (35) Teodoro PE, Teodoro LPR, Baio FHR, da Silva Junior CA, dos Santos RG, Ramos APM, Pinheiro MMF, Osco LP, Gonçalves WN, Carneiro AM, Junior JM, Pistori H, Shiratsuchi LS. Predicting days to maturity, plant height, and grain yield in soybean: a machine and deep learning approach using multispectral data. Remote Sens. 2021;13(22):4632. Doi: 10.3390/rs13224632.



- (36) Volpato L, Dobbels A, Borem A, Lorenz AJ. Optimization of temporal UAS-based imagery analysis to estimate plant maturity date for soybean breeding. Plant Phenome J. 2021;4(1):e20018. Doi: 10.1002/ppj2.20018.
- (37) Hu J, Yue J, Xu X, Han S, Sun T, Liu Y, Feng H, Qiao H. UAV-based remote sensing for soybean FVC, LCC, and maturity monitoring. Agriculture. 2023;13(3):692. Doi: 10.3390/agriculture13030692.
- (38) Zhang S, Feng H, Han S, Shi Z, Xu H, Liu Y, Feng H, Zhou C, Yue J. Monitoring of soybean maturity using UAV remote sensing and deep learning. Agriculture. 2023;13(1):110. Doi: 10.3390/agriculture13010110.
- (39) Pérez O, Diers B, Martin N. Maturity prediction in soybean breeding using aerial images and the Random forest machine learning algorithm. Remote Sens. 2024;16(23):4343. Doi: 10.3390/rs16234343.
- (40) Jiang Y, Li C. Convolutional neural networks for image-based high-throughput plant phenotyping: a review. Plant Phenomics. 2020;2020:4152816. Doi: 10.34133/2020/4152816.
- (41) Zhang Z, Flores P, Igathinathane C, Naik DL, Kiran R, Ransom JK. Wheat lodging detection from UAS imagery using machine learning algorithms. Remote Sens. 2020;12(11):1838. Doi: 10.3390/rs12111838.
- (42) Breiman L. Bagging predictors. Mach Learn. 1996;24(2):123-40. Doi: 10.1007/BF00058655.
- (43) Breiman L. Random forests. Mach Learn. 2001;45(1):5-32. Doi: 10.1023/A:1010933404324.
- (44) Lee, T-H, Ullah A, Wang R. Bootstrap aggregating and Random forest. In: Fuleky P, editor. Macroeconomic forecasting in the era of big data. Cham: Springer; 2020. p. 389-429. Doi: 10.1007/978-3-030-31150-6_13.
- (45) Bruce RW, Rajcan I, Sulik J. Classification of soybean pubescence from multispectral aerial imagery. Plant Phenomics. 2021;2021:9806201. Doi: 10.34133/2021/9806201.
- (46) Palmer RG, Pfeiffer TW, Buss GR, Kilen TC. Qualitative genetics. In: Roger Boerma H, Specht JE, editors. Soybeans: improvement, production, and uses. 3rd ed. Madison: American Society of Agronomy; 2004. p. 137-233. Doi: 10.2134/agronmonogr16.3ed.c5.
- (47) Yan F, Githiri SM, Liu Y, Sang Y, Wang Q, Takahashi R. Loss-of-function mutation of soybean R2R3 MYB transcription factor dilutes tawny pubescence color. Front Plant Sci. 2020;10:1-12. Doi: 10.3389/fpls.2019.01809.
- (48) Roth L, Barendregt C, Bétrix C-A, Hund A, Walter A. High-throughput field phenotyping of soybean: spotting an ideotype. Remote Sens Environ. 2022;269:112797. Doi: 10.1016/j.rse.2021.112797.
- (49) Sarkar S, Zhou J, Scaboo A, Zhou J, Aloysius N, Lim TT. Assessment of soybean lodging using UAV imagery and machine learning. Plants. 2023;12(16):2893. Doi: 10.3390/plants12162893.
- (50) Eftekhari M, Ma C, Orlov YL. Applications of artificial intelligence, machine learning, and deep learning in plant breeding. Front Plant Sci. 2024;15:1-4. Doi: 10.3389/fpls.2024.1420938.
- (51) Abd-Elrahman A, Guan Z, Dalid C, Whitaker V, Britt K, Wilkinson B, Gonzalez A. Automated canopy delineation and size metrics extraction for strawberry dry weight modeling using raster analysis of high-resolution imagery. Remote Sens. 2020;12(21):1-17. Doi: 10.3390/rs12213632.
- (52) Krause MR, Mondal S, Crossa J, Singh RP, Pinto F, Haghighattalab A, Shrestha S, Rutkoski J, Gore MA, Sorrells ME, Poland J. Aerial high-throughput phenotyping enables indirect selection for grain yield at the early generation, seed-limited stages in breeding programs. Crop Sci. 2020;60(6):3096-114. Doi: 10.1002/csc2.20259.
- (53) Maimaitijiang M, Sagan V, Sidike P, Maimaitiyiming M, Hartling S, Peterson KT, Maw MJW, Shakoor N, Mockler T, Fritschi FB. Vegetation index weighted canopy volume model (CVM_{VI}) for soybean biomass estimation from unmanned aerial system-based RGB imagery. ISPRS J Photogramm Remote Sens. 2019;151:27-41. Doi: 10.1016/j.isprsjprs.2019.03.003.
- (54) Walter J, Edwards J, McDonald G, Kuchel H. Photogrammetry for the estimation of wheat biomass and harvest index. Field Crops Res. 2018;216:165-74. Doi: 10.1016/j.fcr.2017.11.024.

- (55) Maimaitijiang M, Sagan V, Sidike P, Hartling S, Esposito F, Fritschi FB. Soybean yield prediction from UAV using multimodal data fusion and deep learning. Remote Sens Environ. 2020;237:111599. Doi: 10.1016/j.rse.2019.111599.
- (56) Hatton NM, Menke E, Sharda A, van der Merwe D, Schapaugh W. Assessment of sudden death syndrome in soybean through multispectral broadband remote sensing aboard small unmanned aerial systems. Comput Electron Agric. 2019;167:105094. Doi: 10.1016/j.compag.2019.105094.
- (57) Marston ZPD, Cira TM, Hodgson EW, Knight JF, Macrae IV, Koch RL. Detection of stress induced by soybean aphid (Hemiptera: Aphididae) using multispectral imagery from unmanned aerial vehicles. J Econ Entomol. 2020;113(2):779-86. Doi: 10.1093/jee/toz306.
- (58) Zhang T, Guan H, Ma X, Shen P. Drought recognition based on feature extraction of multispectral images for the soybean canopy. Ecol Inf. 2023;77:102248. Doi: 10.1016/j.ecoinf.2023.102248.
- (59) Poley LG, McDermid GJ. A systematic review of the factors influencing the estimation of vegetation aboveground biomass using unmanned aerial systems. Remote Sens. 2020;12(7):1052. Doi: 10.3390/rs12071052.
- (60) Benos L, Tagarakis AC, Dolias G, Berruto R, Kateris D, Bochtis D. Machine learning in agriculture: a comprehensive updated review. Sensors. 2021;21(11):3758. Doi: 10.3390/s21113758.